



HEMATOLOGY, TRANSFUSION AND CELL THERAPY

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Images in Clinical Hematology

VEXAS syndrome: more than just vacuoles

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ARTICLE INFO

Article history:

Received 22 May 2025

Accepted 8 June 2025

Available online xxx

1 A 78-year-old man with an autoimmune disorder (vasculitis)
 2 and a recent diagnosis of VEXAS syndrome, confirmed by
 3 next generation sequencing (presence of the p.Met41Thr vari-
 4 ant of the UBA1 gene with a variant allele frequency of 56.4 %)
 5 was admitted to hospital.

6 Analytically he presented: erythrocytes $3.75 \times 10^{12}/L$
 7 (reference values [RV]: $4.5\text{--}5.9 \times 10^{12}/L$), hemoglobin 132 g/L
 8 (RV: 130–175 g/L), mean corpuscular volume 103.6 fL
 9 (RV: 80–97 fL), leucocytes $3.9 \times 10^9/L$ (69 % neutrophils,
 10 20 % lymphocytes), and platelets $197 \times 10^9/L$ (RV: 150–
 11 $450 \times 10^9/L$).

12 The bone marrow aspirate smears were normocellular,
 13 with a myeloid to erythroid (M:E) ratio of 2:1. The smears also
 14 revealed 20 % myeloid precursors (all stages) and 30 % proery-
 15 throblasts with cytoplasmic vacuoles. Other notable findings
 16 included pseudo-Pelger-Huet anomalies and megaloblastic
 17 precursors. Megakaryocytes exhibited a high nuclear-to-cyto-
 18 plasmic ratio; specifically, 50 % of total megakaryocytes (TM)
 19 were sometimes monolobated with eccentrically placed
 20 nuclei, or showed a wreath-like rearrangement of nuclear
 21 lobes. Additionally, multinuclear megakaryocytes (3 % TM)
 22 and megakaryocyte emperipoiesis (3 % TM) were observed
 23 (Figure 1). There was no increase in blasts. Storage iron was
 24 decreased with no ring sideroblasts.

VEXAS syndrome (vacuoles, E1 enzyme, X-linked, autoin- 25
 26 flammatory, somatic) was first reported by Beck et al. in 2020.
 27 Vacuoles are observed in erythroid and myeloid precursor
 28 cells [1]. The E1 enzyme is related to the ubiquitin activating
 29 enzyme encoded by the UBA1 gene, an X-linked gene [1].
 30 Mutation in this gene is responsible for an autoinflammatory
 31 disease (characterized by recurrent fevers, cytopenias, chon-
 32 dritis, vasculitis, pulmonary inflammation, and neutrophilic
 33 dermatoses) as the result of somatic mutations in the
 34 blood [1].

The most frequent mutations are p.Met41Thr (49 %), p. 35
 Met41Val (26 %) and p.Met41Leu (19 %) [2]. 36

In VEXAS syndrome the bone marrow is usually hypercel- 37
 38 lular with an increased M:E ratio ($>4:1$ in $>70\%$ of cases) [2].
 39 The presence of $\geq 10\%$ of myeloid precursors with >1 vacuole
 40 can be both sensitive and specific for VEXAS syndrome, [3]
 41 however cytoplasmic vacuolization of myeloid and erythroid
 42 precursors can be found in other clinical settings: alcohol
 43 abuse, copper deficiency, treatments (chemotherapy and
 44 antibiotics), zinc toxicity, myelodysplastic syndrome, lym-
 45 phoproliferative disorders, multiple myeloma, myeloprolifer-
 46 ative neoplasms and acute myeloid leukemia, or as an
 47 artifact of sample preparation [2,3]. Furthermore, some atyp-
 48 ical UBA1 variants can present absence of precursor vacuoliza-
 49 tion in the bone marrow [2].

Storage iron is usually increased with no significant num- 50
 51 ber of ring sideroblasts ($<10\%$ of cases) [2].

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<https://doi.org/10.1016/j.htct.2025.103984>

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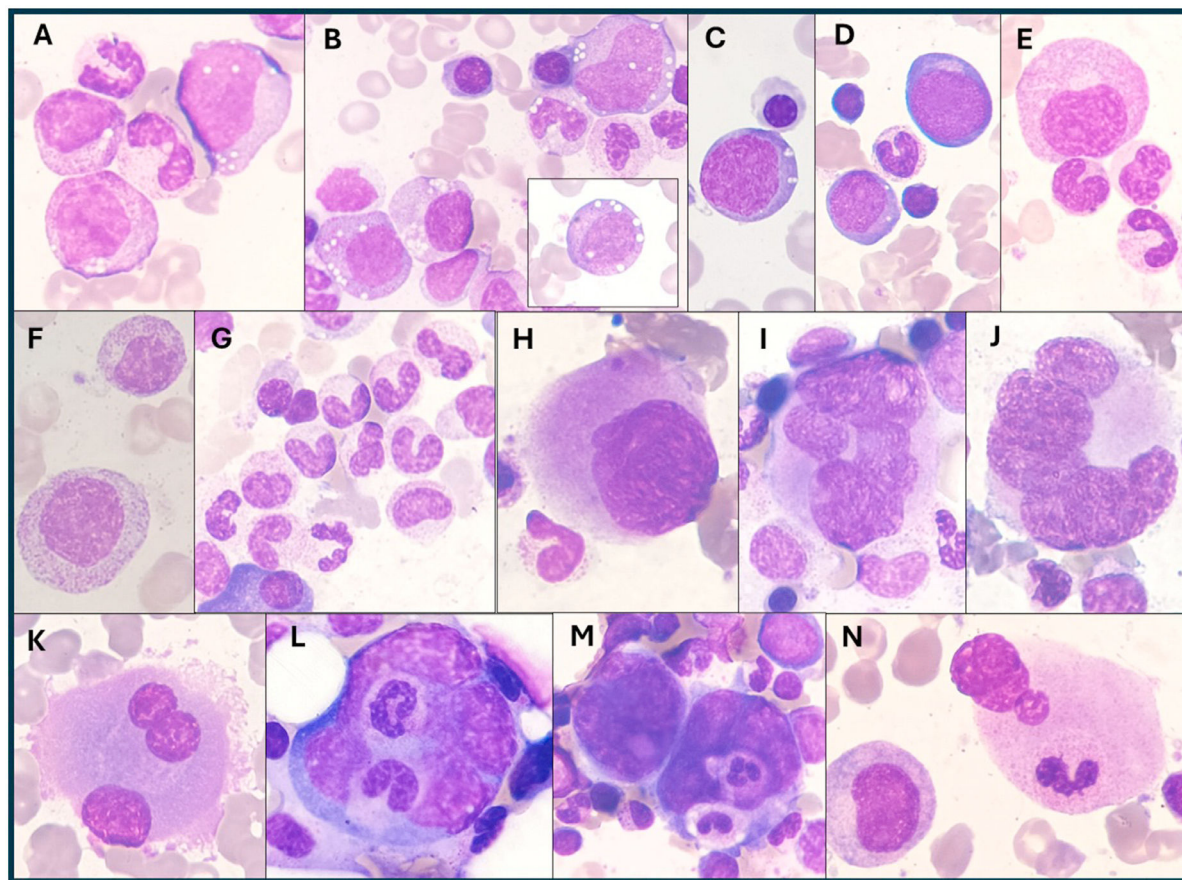


Figure 1 – Bone marrow aspirate (May-Grünwald-Giemsa stain, × 100 objective): Myeloid precursors with vacuoles (A, B); proerythroblasts with vacuoles (C, D); megaloblastic precursors (D, E, F); pseudo-Pelger-Hüet (G); monolobated megakaryocytes (H); megakaryocytes with a wreath-like rearrangement of nuclear lobes (I, J); multinuclear megakaryocytes (K); megakaryocytes emperipolesis (L, M, N).

The full blood count can show: macrocytic anemia (90–100 %), lymphopenia (60–80 %), monocytopenia (50 %), neutropenia (<30 %) and thrombocytopenia (45–69 %) [2].

The UBA1 gene mutation is also a predisposing factor for myelodysplastic syndromes, plasma cell proliferation disorders (monoclonal gammopathy of undetermined significance, multiple myeloma) or both [2].

Conflicts of interest

The author declares no conflicts of interest.

REFERENCES

- Grayson PC, Patel BA, Young NS. VEXAS syndrome. *Blood*. 2021;137(26):3591–4. <https://doi.org/10.1182/blood.2021011455>.
- Koster MJ, Lasho TL, Olteanu H, et al. VEXAS syndrome: clinical, hematologic features and a practical approach to diagnosis and management. *Am J Hematol*. 2024;99(2):284–99. <https://doi.org/10.1002/ajh.27156>.
- Cherniawsky H, Friedmann J, Nicolson H, et al. VEXAS syndrome: a review of bone marrow aspirate and biopsies reporting myeloid and erythroid precursor vacuolation. *Eur J Haematol*. 2023;110(6):633–8. <https://doi.org/10.1111/ejh.13944>.