



Special article

Associação Brasileira de Hematologia, Hemoterapia e Terapia Celular Consensus on genetically modified cells. I: Structuring centers for the multidisciplinary clinical administration and management of CAR-T cell therapy patients



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ABSTRACT

Chimeric antigen receptor T-cells (CAR-T cells) are a new modality of oncological treatment which has demonstrated impressive response in refractory or relapsed diseases, such as acute lymphoblastic leukemia (ALL), lymphomas, and myeloma but is also associated with unique and potentially life-threatening toxicities. The most common adverse events (AEs) include cytokine release syndrome (CRS), neurological toxicities, such as the immune effector cell-associated neurotoxicity syndrome (ICANS), cytopenias, infections, and hypogammaglobulinemia. These may be severe and require admission of the patient to an intensive care unit. However, these AEs are manageable when recognized early and treated by a duly trained team. The objective of this article is to report a consensus compiled by specialists in the fields of oncohematology, bone marrow transplantation, and cellular therapy describing recommendations on the Clinical Centers preparation, training of teams that

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will use CAR-T cells, and leading clinical questions as to their use and the management of potential complications.

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Introduction

The advent of the chimeric antigen receptor T-cell (CAR-T) has revolutionized how we treat hematological neoplasms. Patients with relapsed or refractory diseases, formerly with limited therapeutic options and poor prognosis, began having an effective alternative, with impressive and long-lasting response rates and, in some cases, an improvement in overall survival.^{1–3} However, this new therapeutic modality is associated with unique toxicities, many unknown to hematologist physicians and potentially severe or even fatal.⁴ The early recognition and adequate treatment of the potential complications related to the use of CAR-T cells are imperative for their success.

In this manner, considering imminent approval of some commercial CAR-T products by the *Agência Nacional de Vigilância Sanitária* (ANVISA), the *Associação Brasileira de Hematologia, Hemoterapia e Terapia Celular* (ABHH) invited a panel of specialists in oncohematology diseases, cellular therapy, and bone marrow transplantation to elaborate recommendations on this innovative therapeutic modality to Clinical Centers which will make use of it.

In this article, we describe practical recommendations for preparing Clinical Centers willing to perform CAR-T cell treatment, for the patient care before, during, and following the administration of the cellular product, and, mainly, for the recognition and management of CAR-T related toxicities. Issues related to cell-processing, regulatory and treatment indication are covered in other specific chapters of this Consensus.

Preparation of the center to perform the clinical use of CAR-T cells

The administration of CAR-T cells should be performed at a clinical care unit with structure and expertise in cytotoxic and immunosuppressive drug use and infusion of cryopreserved or fresh cellular products.⁵ A trained team to promptly recognize and treat any acute or late complication due to CAR-T cells use must be available during patient hospitalization, as well as for outpatient evaluations following the discharge from the hospital.

Periodic meetings with the team composed of hematological oncologists, specialists in bone marrow transplant, and cellular therapy are recommended for the evaluation and discussion on the indication for patient inclusion in the treatment.^{6,7} The Center willing to perform the treatment with CAR-T cells must initially be prepared for a high demand for the service and respond with urgency, given the frequent

gravity of the patient condition and absence of other alternative therapies.⁷ The development of a structure which facilitates and organizes the referral of outpatients with a rapid response and the establishment of well-defined criteria for eligibility is the first step before initiating and divulging the treatment. Subsequent chapters of this Consensus may assist in these definitions, which will be fundamental for offering this treatment modality to those indicated.

In Brazil, there are yet no specific regulations for Centers that will make clinical use of CAR-T. However, the regulatory entities may require the participating Centers to be included and trained in risk evaluation and mitigation strategies (REMS) programs similar to those required by the Food and Drug Administration (FDA).^{8,9}

Recommendations

So that a Center may be apt to perform the clinical use of CAR-T cells, the following minimum requirements are recommended:

- Team of hematologist physicians trained to recognize and treat CAR-T complications available 24 h / 7 days;
- The adequate proportion of at least one nurse for four patients in the unit where the treatment will be administered, guaranteeing the continuous monitoring of a patient;
- Available and nearby Intensive Care Unit (ICU), with collaboration well established among the sectors and intensivist professionals familiarized with this treatment modality and its toxicities;
- Team of neurologist physicians trained to recognize and treat neurological complications, such as Immune Effector Cell-Associated Neurotoxicity Syndrome (ICANS);
- Emergency unit with a trained team to recognize and treat CAR-T complications available following hospital discharge;
- Transfusion service;
- Immediate availability of tocilizumab (at least two doses before each treatment) at the unit where the treatment will be administered, and;
- All the members of the teams described above should be trained to recognize and treat all potential complications stemming from CAR-T cell therapy. Periodic meetings among groups and the elaboration of a standard operating procedures manual (SOPM) are further recommended.

These prerequisites are all (except the last) requirements for the accreditation and qualification of a bone marrow transplant (BMT) Center, either autologous or allogeneic, in Brazil.¹⁰ In this manner, this Consensus recommends that CAR-T cell treatment be performed at Centers having allogeneic or autologous BMT service already established, as in the

majority of Centers in Europe and the USA.¹¹ BMT and/or cellular therapy accreditation is also stimulated.

Patient selection and pre-treatment evaluation

The selection of eligible patients is primordial to the success and safety of the CAR-T cell treatment.^{12–14} Initially, it is imperative to guarantee that the patient may benefit from the treatment, selecting only carriers of pathologies for which the specific type of CAR-T cells available has already had its efficacy proven in previous clinical studies. Moreover, to mitigate therapy-associated risks, selection of patients with good performance status, without organic dysfunctions or comorbidities which potentially compromise the capacity to tolerate toxicities related to the treatment is mandatory.¹³ Most of the clinical studies on CAR-T cells exclude patients with organic dysfunction or poor performance status.¹² Although real-life studies have recently demonstrated the safety of using CAR-T cells in older populations, with comorbidities and worse performance status,¹⁵ this conduct is not initially recommended before the Brazilian Centers gain more experience, and there has been a more extended follow-up in these studies to evaluate the risk/benefit ratio.^{12,13}

The initial evaluation should include cardiac, renal, hepatic, and pulmonary function tests and basal neurological examination, similar to the autologous or allogeneic BMT. Furthermore, an active and non-controlled infection may be aggravated by the lymphodepletion caused by chemotherapy, B cell aplasia induced by CAR-T (in anti-CD19 products), or even increase acute toxicities due to elevated basal levels of interleukins. Basal serological evaluation is also recommended, as for autologous or allogeneic BMT.¹²

Active autoimmune diseases can increase the toxicity of CAR-T cells treatment. Moreover, immunosuppressives can interfere in T cells collection, CAR-T cell function, and cellular manufacturing process.¹³

There is a variable and considerable time lapse between patient inclusion and the hospitalization to receive treatment (time necessary for cellular product manufacturing). It is recommended that this same evaluation be repeated before initiating the lymphodepletion regime.

Recommendations

- CAR-T cells indications should be approved on the commercial product label (type of disease, age of the patient, and treatment line). In clinical trials, must respect the inclusion and exclusion criteria;
- Patients with a good performance status (ECOG 0-2) and without severe organic dysfunctions should be eligible for the treatment with CAR-T cells;
- Treatment with CAR-T cells is not recommended for patients with active autoimmune disease resulting in end-organ injury or requiring systemic immunosuppression;
- Patients with non-controlled active infection should be adequately treated, and the CAR-T cell therapy postponed until its resolution or control;

- Tests recommended for initial evaluation (eligibility), and those before the initiation of lymphodepletion are described in Table 1, and;
- We recommend that an informed consent form (ICF) be obtained from the patient and/or its guardian before any procedure, similar to what already occurs for BMT.

Conditioning or lymphodepletion

Conditioning with lymphodepletion chemotherapeutic schemes before CAR-T cells infusion increases the efficacy of the treatment.¹⁶ Multiple mechanisms, such as immune suppressor cells elimination (regulatory T-cells, for example), reduction of T, B, and NK cells, as well as an increase in homeostatic cytokines (IL-7 and IL-15), and an increase in tumor cell co-stimulatory molecules promote a favorable environment for the expansion and survival of *in vivo* CAR-T cells.¹⁷

Fludarabine and cyclophosphamide combination was superior to cyclophosphamide alone or cyclophosphamide plus etoposide¹⁸ and is currently more widely used and recommended. However, various other schemes containing bendamustine, pentostatin, total body irradiation (TBI), and polychemotherapy, although possible, are seldom used.^{12–14}

Recommendations

- Initiate conditioning only following the confirmation of the availability of the cellular product (CAR-T cells) for the patient, considering the product estimated date of arrival at the Center;
- Conditioning should be performed in all patients, independent of previous leukocyte or lymphocyte counts;
- Fludarabine and cyclophosphamide regimen is recommended for conditioning; and
- CAR-T cells may cause rapid destruction of tumor cells. Tumor lysis prophylaxis is recommended, as per risk and institutional protocol, considering the type of disease, tumor load, and comorbidities.

Infusion of CAR-T cells

The infusion of cellular products is generally safe and well-tolerated, although severe adverse reactions may occur and should be monitored. For such, the patient must be hospitalized and monitored for vital signs, oxygen saturation, and urinary output.^{12,14} Emergency materials, aspiration, and oxygen should be available at the patient's bedside.

Characteristic reactions, such as nausea, vomiting, abdominal pain, fever, and tremors, are the most frequent. Rarely, respiratory depression, cardiac arrhythmias, and neurological symptoms may be observed.¹⁹ The general principles for managing these reactions include reducing the velocity or halting of the infusion, standard emergency conduct, and the confirmation of the receptor/product.

Table 1 – Recommended minimum evaluation for determination of eligibility to CAR-T cells clinical use.

Evaluation	Comment
General physical examination	Baseline.
Neurological examination	Baseline.
Disease confirmation (refractoriness/relapse)	Histological for Lymphoma/Multiple Myeloma. Immunophenotyping for Acute Lymphoid Leukemia.
Echocardiogram	Requires LVEF > 40%.
ECG	Baseline.
Creatinine clearance	Requires > 30 mL/min. Be careful with patients with clearance < 60 mL/min.
Electrolytes (Sodium, Potassium, Calcium, Phosphorus)	Apheresis for the lymphocyte collection may precipitate hydroelectrolytic disturbances.
Uric Acid	For evaluation of Tumor Lysis Syndrome risk.
LDH	For evaluation of Tumor Lysis Syndrome risk.
ALT/AST	For evaluation of tumor load and tumor Lysis Syndrome risk.
Bilirubin	Recommended < 5 x LSN (except if attributed to hepatic infiltration by baseline disease).
Blood Count	Recommended BT < 2 mg/dL.
C-reactive Protein	Recommended Neutrophils > 1000/ mm ³ .
HIV	Recommended for evaluating active infection.
Hepatitis B	Positive serology contraindicates cellular collection and processing by some suppliers of commercial CAR-T products. Prophylaxis against the hepatitis B virus is recommended for patients with positive serology indicating infection, with or without viral replication, for a minimum period of 6 months. Initiate before conditioning.
Hepatitis C	Positive serology contraindicates cellular collection and processing by some suppliers of commercial CAR-T products.
Serology for Chagas	Positive serology contraindicates cellular collection and processing by some suppliers of commercial CAR-T products.
Serology for Cytomegalovirus	Do not contraindicate the procedure. Monitor.
HTLV-1	Do not contraindicate the procedure. Monitor.
Serological test for syphilis	Do not contraindicate the procedure. Perform treatment before CAR-T use.
Immunoglobulin level	Do not contraindicate the procedure. Monitor.
Quantification of CD4	Baseline.
CNS Image (NMR)	Baseline.
Lumbar puncture	Obligatory if previous CNS disease or current neurological symptoms. However, it is recommended that all patients perform baseline NMR before CAR-T treatment.
Pregnancy test	Only if previous CNS disease or current neurological symptoms.
	Serum or urinary. It should be negative before leukapheresis and before lymphodepletion.

LVEF: Left ventricular ejection fraction; ECG: Electrocardiogram; LDH: Lactate dehydrogenase; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; HIV: Human immunodeficiency virus; HTLV-1: Human T-lymphotropic virus type 1; CNS: Central nervous system; NMR: Nuclear Magnetic Resonance.

Adapted from: Yakoub-Agha I, et al. *Haematologica*, 2020¹².

Main acute complications generally occur within 28 days of the CAR-T cells infusion and frequently after hospital discharge.¹² Thus, the patient should remain near the Center responsible for the CAR-T cells clinical use, having prompt access to emergency support service during this period.

Recommendations

- CAR-T cells should be infused with the patient hospitalized, with a minimum intrahospital observation for 14 days. This recommendation stems from the Centers gaining experience with this treatment modality;
- There should be a two-person (double-check) verification of the product data and confirmation of the receptor identification before installing the cells;
- Must use connections or transfusion equipment without a leukocyte filter;

- Paracetamol and an antihistamine (diphenhydramine) should be used as pre-medication, 30 to 60 minutes before cell infusion;
- Corticosteroids should not be used previously to or immediately after CAR-T cells infusion (except in cases of anaphylactic reaction);
- No medication should be infused in the same venous line concomitantly with CAR-T cells;
- A rapid infusion is recommended, in approximately 30 min;
- The patient should be monitored during the whole procedure and in the subsequent hours;
- The preparation and supply of materials enlightening patients on the treatment and possible complications is recommended. A pocket card with orientations on the alert signs/symptoms and contact information of the Center responsible for CAR-T clinical use should be in the patient's possession following the hospital discharge,^{8,9} and;

- The patient should remain close to the Center responsible for CAR-T clinical use (up to two hours away is recommended) for at least four weeks following the infusion.

Management of CAR-T cells complications

The two most frequent CAR-T cell-related complications are cytokine release syndrome (CRS) and ICANS. These toxicities can be severe or even fatal, and early recognition is crucial for their management. CRS and ICANS occur days to weeks following CAR-T cells infusion (depending on the product infused), when most of the patients also have post-chemotherapy pancytopenia, and associated infections may complicate the diagnosis and treatment of these syndromes.

Projects with CAR-T cells are under development in Brazilian research centers, and different products are being approved for commercial use in Brazil. The clinical presentation of CRS and ICANS is similar for the various products; however, the initiation time, incidence, and severity can significantly vary. Products' particularities, such as the type of cells used in the production (mononuclear cells vs. T cells), differences in the fabrication process, co-stimulatory molecule (CD28 vs. 4-1BB), CAR-T cells dose, and the type and intensity of the lymphodepletion chemotherapy, as well as patient-specific factors, such as age, diagnosis, and tumor load, may influence in these differences. Clinical studies with anti-CD19 CAR-T cells containing CD28 as the co-stimulatory molecule (e.g., axicabtagene ciloleucel) suggest greater risk and severity of CRS and ICANS when compared to products containing 4-1BB.

Considering these particularities, this Consensus presents recommendations for CRS and ICANS management, primarily in adults. Management of CRS and ICANS is based on the American Society for Transplantation and Cellular Therapy (ASTCT) classification system.²⁰ For commercial products, additional recommendations present in the package insert and in the REMS should also be taken into consideration.^{8,9}

Cytokine Release Syndrome (CRS)

CRS presents as a systemic inflammatory response syndrome with fever (not attributed to any other cause). It is fundamental to evaluate, treat and prevent systemic infection, as most patients present with febrile neutropenia and various CRS manifestations mimic sepsis. Moreover, infectious conditions and systemic inflammation can exacerbate the CRS. In this manner, both possibilities should be considered and treated in a patient with fever within the first weeks following CAR-T cell therapy.

Table 2 presents the main symptoms and laboratory findings of CRS. Following CRS diagnosis, the persistence of fever is not necessary for CRS classification.

It is recommended to evaluate vital signs every 4 h following CAR-T cell infusion, and daily blood count, renal function, electrolytes, hepatic function, lactate dehydrogenase, coagulation tests (including fibrinogen and d-dimers), uric acid, c-reactive protein (CRP), and ferritin.

Table 2 – Symptoms and laboratory findings in CRS.

Symptoms	
Constitutional	Fever, tremors, fatigue, anorexia, myalgia, arthralgia, headache
Dermatological	Rash
Gastrointestinal	Nausea, vomiting, diarrhea
Respiratory	Tachypnea, hypoxemia
Cardiovascular	Tachycardia, hypotension, thoracic pain, arrhythmias
Hematological	Hemorrhage, disseminated intravascular coagulation
Renal	Acute renal insufficiency
Laboratory findings	
Hematological	D-dimers elevation, hypofibrinogenemia, PT, and PTT prolongation
Renal	Hyponatremia, hypokalemia, hypophosphatemia
Hepatic	Elevated transaminases, hyperbilirubinemia
Others	Elevated C-reactive protein, ferritin, and interleukin-6

Table 3 presents recommendations for the management based on the ASTCT classification system. CRS treatment is based on the use of tocilizumab (anti-interleukin-6 receptor (IL-6) antibody) or other anti-IL-6 therapy available/approved in the country. The hospitals/centers should maintain at least two anti-IL-6 doses available for administration in up to two hours.

In the Brazilian Centers' initial experience with CAR-T cells, it will be important that intensivist physicians be aware of the CAR-T cell infusions and follow CRS management, even in low-grade cases, so that there will be greater continuity in the treatment of these patients. The same should occur with neurologists and infectious disease specialists physicians.

Cell-Associated Neurotoxicity Syndrome (ICANS)

Neurotoxicity mainly occurs in the first 1–2 weeks following CAR-T cell infusion and rarely later on. ICANS evaluation involves using the Immune Effector Cell Encephalopathy (ICE) scale, with a 0–10 point system (Table 4). Patients should be submitted to a basal neurological evaluation, including the ICE scale, with daily reevaluation from infusion until the 21st day following the administration of the CAR-T cells. Antiepileptic prophylaxis with agents, such as levetiracetam, is not routinely recommended, except for patients with a history of convulsions or central nervous system disease.¹²

Table 5 presents the management recommendations based on the ASTCT classification system. Tocilizumab does not cross the hematoencephalic barrier and is associated with an increase in IL-6 serum levels and potentially greater risk and severity of ICANS due to the rise in IL-6 levels in the central nervous system. For this reason, tocilizumab should not be used for the treatment of ICANS in the absence of CRS.

Infections

Infections related to CAR-T cell therapy are prevalent events. Febrile neutropenia grade ≥ 3 occurs in approximately 15 to

Table 3 – CRS classification and management.

Grade	Definition	Management
1	<ul style="list-style-type: none"> • Temperature $\geq 38^{\circ}\text{C}^*$ 	<ul style="list-style-type: none"> • Symptomatic. • Evaluation, prophylactic and empirical treatment of infections in neutropenic patients. • Consider tocilizumab 8 mg/kg IV (maximum 800 mg) single dose in patients with fever in the first 24 h following CAR-T cell infusion and/or with high tumor load. • Consider initiating oral prophylactic levetiracetam (500 mg every 12 h).
2	<ul style="list-style-type: none"> • Temperature $\geq 38^{\circ}\text{C}^*$ • Hypotension without vasoactive drug • and/or[†] • Hypoxia requiring $\text{O}_2 \leq 6\text{L}/\text{min}$ 	<ul style="list-style-type: none"> • Crystalloid solution and additional oxygen bolus. • Evaluation, prophylactic and empirical treatment of infections in neutropenic patients. • Tocilizumab 8 mg/kg IV (maximum 800 mg) single dose. Repeat in 8–12 h in the absence of response. Dexamethasone 10 mg IV every 12 h can be considered with the first dose and is indicated with the second dose of tocilizumab. • In the absence of response following two doses of tocilizumab and dexamethasone, third-line agents (anakinra, siltuximab, and pulse methylprednisolone) should be considered. • Initiate oral prophylactic levetiracetam (500mg every 12 h). • Telemetry monitoring.
3	<ul style="list-style-type: none"> • Temperature $\geq 38^{\circ}\text{C}^*$ • Hypotension requiring vasoactive drug with or without vasopressin • and/or[†] • Hypoxia requiring $\text{O}_2 > 6\text{L}/\text{min}$ 	<ul style="list-style-type: none"> • Transfer to intensive care unit (ICU). • Evaluation, prophylactic and empirical treatment of infections in neutropenic patients. • Tocilizumab 8 mg/kg IV (maximum 800 mg) single dose. Repeat in 8–12 h in the absence of response (maximum of 3 doses in 24 h). • Dexamethasone 10 mg IV every 6 h. • In the absence of response following two doses of tocilizumab and dexamethasone, third-line agents (anakinra, siltuximab, and pulse methylprednisolone) should be considered. • Initiate oral prophylactic levetiracetam (500 mg every 12 hours). • Telemetry monitoring.
4	<ul style="list-style-type: none"> • Temperature $\geq 38^{\circ}\text{C}^*$ • Hypotension requiring multiple vasoactive drugs (excluding vasopressin) • and/or[†] • Hypoxia requiring positive pressure 	<ul style="list-style-type: none"> • Transfer to ICU/ remain in ICU. • Evaluation, prophylactic and empirical treatment of infections in neutropenic patients. • Tocilizumab 8 mg/kg IV (maximum 800 mg) single dose. Repeat in 8–12 h in the absence of response (maximum three doses in 24 h). • Dexamethasone 10 mg IV every 6 h. Consider pulse methylprednisolone 1g IV 1 time/day for three days if there is instability, lack of improvement, or worsening. • In the absence of response following two doses of tocilizumab and corticosteroid, third-line agents (anakinra, siltuximab, and pulse methylprednisolone) should be considered. • In patients without improvement or with rapid worsening following the introduction of third-line agents and/or findings suggestive of macrophage activation syndrome/hemophagocytic syndrome, consider cytotoxic agents (e.g., etoposide). • Initiate oral prophylactic levetiracetam (500mg every 12 h). • Telemetry monitoring.

Modified from Lee DW, et al. Biol Blood Marrow Transplant. 2019.²⁰

* Fever with no other cause. In CRS patients who receive antipyretics or anti-cytokine therapy, such as tocilizumab and corticosteroids, persistent fever is not necessary for the CRS classification. In this case, the classification is based on hypotension and/or hypoxia.

[†] The CRS grade is determined by the gravest event: hypotension or hypoxia with no other cause.

Table 4 – ICE scale.

Domain	Points
Orientation: year, month, city, hospital	4
Nomination: 3 objects	3
Follow-up: simple commands	1
Writing: standard sentence	1
Attention: regressive count from 100 subtracting 10	1

Modified from Lee DW, et al. Biol Blood Marrow Transplant. 2019.²⁰

35% of cases in the first four to eight weeks following infusion.^{12,21–24} During the first weeks, bacteremia, fungal infections, upper respiratory infections, herpes zoster, and viral reactivations, such as cytomegalovirus, Epstein Barr virus, and human herpesvirus-6 may appear²⁵; and after 30 days, viral respiratory infections, cytomegalovirus, and pneumonia predominate.^{12,22,23,26} Invasive fungal infections are rare and

mainly observed in ALL patients who have previously had allogeneic BMT.¹²

Described risk factors for CAR-T-related infection are more than four previous chemotherapy schemes, infusion of high CAR-T dose, CRS grade ≥ 3 , use of systemic corticosteroid, and serum IgG $< 400\text{ mg}/\text{dL}$.^{23,26–29}

Recommendations

- Routine antibacterial prophylaxis is not recommended;
- Use viral prophylaxis with acyclovir 400 mg every 12 h (Pediatric dose: 60–90 mg/kg/day - maximum of 800mg, divided into 2 or 3 doses. Children and Adolescents $\geq 40\text{ kg}$: same dose as for adults) or valaciclovir 500 mg every 12 h (Pediatric dose for patients $< 40\text{ kg}$: 250 mg. Children and Adolescents $\geq 40\text{ kg}$: same dose as for adults)^{12,25}; for 6–12 months following CAR-T infusion, preferentially suspending only after $\text{CD4} > 200/\text{mm}^3$;

Table 5 – Classification and management of ICANS.

Grade	ICE Scale	Consciousness level [†]	Epileptic seizure	Motor findings [‡]	Intracranial hypertension/cerebral edema [§]	Management
1	7–9	Rouses spontaneously	NA	NA	NA	<ul style="list-style-type: none"> • Initiate oral prophylactic levetiracetam (500mg every 12 h). • Electroencephalogram (EEG) and neuroimaging. • Consider tocilizumab if there is concomitant CRS.
2	3–6	Rouses with auditory stimulus	NA	NA	NA	<ul style="list-style-type: none"> • Initiate oral prophylactic levetiracetam (500mg every 12 h). • Dexamethasone 10mg IV every 12 hours. • Consider tocilizumab if there is concomitant CRS.
3	0–2*	Rouses only with tactile stimulus	Any epileptic seizure, focal or generalized with rapid resolution or non-convulsive seizure on EEG which respond to intervention	NA	Focal/ local edema in neuroimage	<ul style="list-style-type: none"> • Consider transfer to the Intensive Care Unit (ICU). • Initiate oral prophylactic levetiracetam (500 mg every 12 h). If there are epileptic seizures, consider the association with other anticonvulsants. • Dexamethasone 10 mg IV every 6–12 h. Consider pulse methylprednisolone 1g IV 1 time/day for 3 days focal/ local edema. • Consider tocilizumab if there is concomitant CRS. • If there is instability, lack of improvement, or worsening in the absence of CRS, consider other cytokine antagonists (e.g., anakinra).
4	0*	Unconscious or requiring vigorous and repetitive tactile stimulation to wake up. Stupor or coma.	Prolonged epileptic seizure (> 5 min) or state of epileptic malaise.	Focal deep muscular weakness	Diffuse cerebral edema in neuroimage; decerebration or decortication; paralysis of VI cranial pair; papilloedema; Cushing triad	<ul style="list-style-type: none"> • Transfer to ICU. • Initiate oral prophylactic levetiracetam (500 mg every 12 h). If already in use, consider association with other anticonvulsants. • Dexamethasone 10 mg IV every 6 hours. Consider pulse methylprednisolone 1 g IV 1 time/day for three days. • Monitor with telemetry. • Consider tocilizumab if there is concomitant CRS. • If there is instability, lack of improvement, or worsening in the absence of CRS, consider other cytokine antagonists (e.g., anakinra).

Modified from Lee DW, et al. *Biol Blood Marrow Transplant.* 2019²⁰.

* Patients with ICE scale 0 can be classified as ICANS grade 3 if conscious and with global aphasia and ICANS grade 4 if unconscious and incapable of performing the ICE scale.

[†] Lowering of level of consciousness not attributed to other causes (e.g., sedatives).

[‡] Tremors and myoclonia do not influence the ICANS classification.

[§] Intracranial hemorrhage with or without associated edema is not considered a manifestation of neurotoxicity and is excluded from the ICANS classification.

^{||} Anakinra (interleukin-1 receptor antagonist) is used subcutaneously at 1–8mg/kg of current weight divided into 2–3 doses over 24 h. Maximum dose of 8mg/kg over 24 h.

- Use prophylaxis for pneumocystis with sulfamethoxazole/trimethoprim 800/160 mg/day (Pediatric dose: 5–10 mg/kg/day of trimethoprim), 3 times per week^{12,25} for 6–12 months following CAR-T infusion, preferentially suspending only after CD4 > 200/mm³;
- Antifungal prophylaxis with fluconazole is recommended for patients using corticosteroid ≥ 0.5 mg/kg/day and during the period of neutropenia. Add filamentous fungi prophylaxis in the presence of two or more of the following risk factors: ≥ 4 previous chemotherapy treatments, neutropenia ($< 500/\text{mm}^3$) before CAR-T infusion, CAR-T dose over $2 \times 10^7/\text{kg}$, previous invasive fungal infection, and use of tocilizumab and/or corticosteroids.²⁹ The choice of the best antifungal agent is at the discretion of each institution, local epidemiology, and previous patient's history;
- Infectious vigilance and adequate antimicrobial prophylaxis are recommended for patients who had received immunosuppressive treatment for CRS and ICANS and patients with prolonged neutropenia using corticosteroids^{12,25};
- Prophylaxis against hepatitis B virus is recommended for patients with positive serology indicating previous infection, with or without viral replication, initiating before the conditioning and maintained for a minimum of 6 months^{12,30};
- Start vaccinations six months after CAR-T cell infusion,³⁰ and;
- Influenza³⁰ and SARS-CoV-2³¹ vaccinations are recommended, completing the scheme two weeks before CAR-T cells infusion.

Cytopenias

Cytopenias are very frequent following anti-CD19 CAR-T cells treatment and are also associated with previous treatments and lymphodepletion before CAR-T infusion. Grade ≥ 3 neutropenia occurs in up to 78% of patients; grade ≥ 3 anemia in up to 43%, and grade ≥ 3 thrombocytopenia in up to 38% of patients.³² The risk factors for CAR-T-related cytopenias are grade ≥ 3 CRS/ICANS, pre-existing cytopenias, and possibly the type of product used (a previous study showed fewer cytopenias following Tisa-Cel).³³ These cytopenias can persist three months after CAR-T infusion in approximately 32% of cases²⁴ and may last even longer.

Levels of CD8+ and CD56+ lymphocytes persist relatively stable following anti-CD19 CAR-T infusion. In contrast, CD4+ lymphocytes remain low, even one year after CAR-T infusion, with a median of approximately 150–250 cells/mm³,^{21,27,34,35} and CD19+ lymphocytes recovery inversely correlates with the persistence of anti-CD19 CAR-T cells.^{36,37}

Recommendations

- Cytopenias lasting over 28 days: proceed with bone marrow aspirate and biopsy to rule out viral infection, myelodysplasia, and disease relapse²⁵;
- G-CSF can be used to reduce the period of neutropenia (if grade ≥ 3).¹² However, some groups advocate G-CSF suspension during the CRS period due to theoretical risk of worsening. GM-CSF is not recommended²⁵;

- Platelets and RBCs transfusion, as per institutional protocol;
- Platelet transfusion refractoriness may occur more frequently during CRS;
- Patients already in outpatient management, with active infections or febrile neutropenia, should be hospitalized and treated according to institutional protocols,²⁵ and;
- Evaluate baseline CD4 and serum IgG levels and at 3, 6, and 12 months following CAR-T infusion.

Hypogammaglobulinemia

Hypogammaglobulinemia is due to the “on-target/off-tumor” effect of anti-CD19 and anti-BCMA CAR-T cells. Almost all patients who respond to anti-CD19 CAR-T therapy develop B-cell aplasia following the infusion,²⁵ which can persist for months to years.¹² Up to 60% of patients present with persistent hypogammaglobulinemia 90 days after the treatment,^{21,22,37} and serum IgG levels appear to reach their nadir approximately six months after CAR-T. In some diseases, 74% of patients may have previous low baseline IgG levels, even before CAR-T therapy. Many of them due to previous anti-CD20 monoclonal antibodies.^{22,38}

Despite the hypogammaglobulinemia, late infections following CAR-T are predominantly caused by respiratory viruses and do not lead to hospitalization. Many patients maintain protective levels of vaccine-induced antibodies and against viruses, despite CD19 and serum IgG levels,^{21,37,39} which appears to be associated with plasmocytes persistence following CD19+ lymphocytes depletion. Children with fewer plasmocytes can be more susceptible to infections than adults,³⁰ and these levels might be lower following anti-BCMA CAR-T.⁴⁰ Pathogens-specific IgG levels were similar for most pathogens in post-CAR-T cell therapy, with and without intravenous immunoglobulin replacement, except for *S. pneumoniae*.⁴⁰ Given these data, costs, and risks associated with intravenous immunoglobulin infusion, we do not recommend its use based solely on serum IgG levels. We suggest intravenous immunoglobulin replacement guided by clinical conditions (ex. patients with severe or recurrent infections and serum IgG < 400 mg/dL). For patients with serum IgG > 400 mg/dL and severe and/or recurrent infections, consider dosage of IgG subclasses and pathogen-specific IgG titers.^{30,37}

Recommendations

- We recommend monitoring B-cells quantification and serum immunoglobulin levels monthly, for at least three months, following CAR-T cells infusion. After six months, monitoring if clinically indicated^{25,41};
- Adults with recurrent or severe infections, and children with serum immunoglobulin level less than 400 mg/dL, should be considered for monthly intravenous immunoglobulin infusion^{25,37};
- The recommended dose is the one that will maintain serum IgG level over 400 mg/dL or normal for the age. Start 400–600mg/kg every 3–4 weeks, and increase dose or frequency of infusions by clinical criteria.²⁹ For children, start 0.5g/kg/monthly.

Conclusion

CAR-T cells therapy is certainly an advance in hematology. It is an effective treatment with a manageable safety profile for some patients with refractory or relapsed oncohematologic diseases. However, this new therapeutic modality also brings further complications and adverse events. The hematologist is not familiar with the CRS and ICANS, which should be expected and promptly approached. Other potential complications, also frequent in regular oncohematologic treatments, such as cytopenias, infections, and hypogammaglobulinemia, present particularities that should be understood. Therefore, it is essential that the multidisciplinary teams that will care for these patients be previously trained and capacitated to recognize and treat these complications; and that the Centers that will perform this type of treatment have an adequate structure. The selection of the patient who will benefit from and tolerate the therapy is another fundamental pillar for the clinical use of CAR-T cells.

Conflicts of interest

The authors declare no conflicts of interest.

REFERENCES

- Quintás-Cardama A. CD19 directed CARs in acute lymphoblastic leukemia: state of the art and beyond. *Leuk Lymphoma*. 2019;60(5):1346–8. <https://doi.org/10.1080/10428194.2018.1533132>.
- Qualls D, Salles G. Optimizing CAR T cell therapy in lymphoma. *Hematol Oncol*. 2021;39(Suppl 1):104–12. <https://doi.org/10.1002/hon.2844>.
- van de Donk NW, Usmani SZ, Yong K. CAR T-cell therapy for multiple myeloma: state of the art and prospects. *Lancet Haematol*. 2021;8(6):e446–61. [https://doi.org/10.1016/S2352-3026\(21\)00057-0](https://doi.org/10.1016/S2352-3026(21)00057-0).
- Brudno JN, Kochenderfer JN. Recent advances in CAR T-cell toxicity: mechanisms, manifestations and management. *Blood Rev*. 2019;34:45–55. <https://doi.org/10.1016/j.blre.2018.11.002>.
- Yakoub-Agha I. Clinical units to set up chimeric antigen receptor T-cell therapy (CAR T-cells): based on the recommendations of the Francophone Society of Bone Marrow Transplantation and Cellular Therapy (SFGM-TC). *Curr Res Transl Med*. 2018;66(2):57–8. <https://doi.org/10.1016/j.retram.2018.04.001>.
- Buechner J, Kersten MJ, Fuchs M, Salmon F, Jäger U. Chimeric antigen receptor-T cell therapy: practical considerations for implementation in Europe. *Hemasphere*. 2018;2(1):e18. <https://doi.org/10.1097/HS9.000000000000018>.
- Perica K, Curran KJ, Brentjens RJ, Giral SA. Building a CAR garage: preparing for the delivery of commercial CAR T cell products at memorial sloan kettering cancer center. *Biol Blood Marrow Transpl*. 2018;24(6):1135–41. <https://doi.org/10.1016/j.bbmt.2018.02.018>.
- Administration. USFaD. Approved risk evaluation and mitigation strategies (REMS). Kymriah (tisagenlecleucel). 2021. Disponível em: <https://www.accessdata.fda.gov/scripts/cder/remis/index.cfm?event=IndvRemsDetails.page&REMS=368>
- Administration. USFaD. Approved risk evaluation and mitigation strategies (REMS). Yescarta and Tecartus (axicabtagene ciloleucel). 2021. Disponível em: <https://www.accessdata.fda.gov/scripts/cder/remis/index.cfm?event=IndvRemsDetails.page&REMS=375>
- BRASIL. Ministério da Saúde. Gabinete do Ministro. Portaria no. 931, de 2 de maio de. Brasília, 2006.
- Hayden PJ, Sirait T, Koster L, Snowden JA, Yakoub-Agha I. An international survey on the management of patients receiving CAR T-cell therapy for haematological malignancies on behalf of the Chronic Malignancies Working Party of EBMT. *Curr Res Transl Med*. 2019;67(3):79–88. <https://doi.org/10.1016/j.retram.2019.05.002>.
- Yakoub-Agha I, Chabannon C, Bader P, Basak GW, Bonig H, Ciceri F, et al. Management of adults and children undergoing chimeric antigen receptor T-cell therapy: best practice recommendations of the European Society for Blood and Marrow Transplantation (EBMT) and the Joint Accreditation Committee of ISCT and EBMT (JACIE). *Haematologica*. 2020;105(2):297–316. <https://doi.org/10.3324/haematol.2019.229781>.
- Jain T, Bar M, Kansagra AJ, Chong EA, Hashmi SK, Neelapu SS, et al. Use of chimeric antigen receptor T cell therapy in clinical practice for relapsed/refractory aggressive B cell on-hodgkin lymphoma: an expert panel opinion from the American Society for transplantation and cellular therapy. *Biol Blood Marrow Transpl*. 2019;25(12):2305–21. <https://doi.org/10.1016/j.bbmt.2019.08.015>.
- Mahadeo KM, Khazal SJ, Abdel-Aziz H, Fitzgerald JC, Taraseviciute A, Bollard CM, et al. Management guidelines for paediatric patients receiving chimeric antigen receptor T cell therapy. *Nat Rev Clin Oncol*. 2019;16(1):45–63. <https://doi.org/10.1038/s41571-018-0075-2>.
- Ghesquieres H, Salles G. Early off-study experience of chimeric antigen receptor T cells in aggressive lymphoma: Closer to a real-world setting. *J Clin Oncol*. 2020;38(27):3085–7. <https://doi.org/10.1200/JCO.20.01134>.
- Hirayama AV, Gauthier J, Hay KA, Voutsinas JM, Wu Q, Gooley T, et al. The response to lymphodepletion impacts PFS in patients with aggressive non-Hodgkin lymphoma treated with CD19 CAR T cells. *Blood*. 2019;133(17):1876–87. <https://doi.org/10.1182/blood-2018-11-887067>.
- Neelapu SS. CAR-T efficacy: Is conditioning the key? *Blood*. 2019;133(17):1799–800. <https://doi.org/10.1182/blood-2019-03-900928>.
- Hay KA, Gauthier J, Hirayama AV, Voutsinas JM, Wu Q, Li D, et al. Factors associated with durable EFS in adult B-cell ALL patients achieving MRD-negative CR after CD19 CAR T-cell therapy. *Blood*. 2019;133(15):1652–63. <https://doi.org/10.1182/blood-2018-11-883710>.
- Shu Z, Heimfeld S, Gao D. Hematopoietic SCT with cryopreserved grafts: adverse reactions after transplantation and cryoprotectant removal before infusion. *Bone Marrow Transpl*. 2014;49(4):469–76. <https://doi.org/10.1038/bmt.2013.152>.
- Lee DW, Santomasso BD, Locke FL, Ghobadi A, Turtle CJ, Brudno JN, et al. ASTCT consensus grading for cytokine release syndrome and neurologic toxicity associated with immune effector cells. *Biol Blood Marrow Transplant*. 2019;25:625–38.
- Logue JM, Zucchetti E, Bachmeier CA, Krivenko GS, Larson V, Ninh D, et al. Immune reconstitution and associated infections following axicabtagene ciloleucel in relapsed or refractory large B-cell lymphoma. *Haematologica*. 2021;106(4):978–86. <https://doi.org/10.3324/haematol.2019.238634>.
- Cordeiro A, Bezerra ED, Hirayama AV, Hill JA, Wu QV, Voutsinas J, et al. Late events after treatment with CD19-targeted chimeric antigen receptor modified T Cells. *Biol Blood Marrow Transpl*. 2020;26(1):26–33. <https://doi.org/10.1016/j.bbmt.2019.08.003>.
- Hill JA, Li D, Hay KA, Green ML, Cherian S, Chen X, et al. Infectious complications of CD19-targeted chimeric antigen receptor-modified T-cell immunotherapy. *Blood*. 2018;131(1):121–30. <https://doi.org/10.1182/blood-2017-07-793760>.

24. Schuster SJ, Bishop MR, Tam CS, Waller EK, Borchmann P, McGuirk JP, et al. Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N Engl J Med*. 2019;380(1):45–56. <https://doi.org/10.1056/nejmoa1804980>.
25. Maus MV, Alexander S, Bishop MR, Brudno JN, Callahan C, Davila ML, et al. Society for Immunotherapy of Cancer (SITC) clinical practice guideline on immune effector cell-related adverse events. *J Immunother Cancer*. 2020;8(2):1–25. <https://doi.org/10.1136/jitc-2020-001511>.
26. Park JH, Rivière I, Gonen M, Wang X, Sénéchal B, Curran KJ, et al. Long-term follow-up of CD19 CAR therapy in acute lymphoblastic leukemia. *N Engl J Med*. 2018;378(5):449–59. <https://doi.org/10.1056/nejmoa1709919>.
27. Wudhikarn K, Palomba ML, Pennisi M, Garcia-Recio M, Flynn JR, Devlin SM, et al. Infection during the first year in patients treated with CD19 CAR T cells for diffuse large B cell lymphoma. *Blood Cancer J*. 2020;10(8). <https://doi.org/10.1038/s41408-020-00346-7>.
28. Vora SB, Waghmare A, Englund JA, Qu P, Gardner RA, Hill JA. Infectious complications following CD19 chimeric antigen receptor t-cell therapy for children, adolescents, and young adults. *Open Forum Infect Dis*. 2020;7(5):1–9. <https://doi.org/10.1093/OFID/OFAA121>.
29. Los-Arcos I, Iacoboni G, Aguilar-Guisado M, Alsina-Manrique L, Díaz de Heredia C, Fortuny-Guasch C, et al. Recommendations for screening, monitoring, prevention, and prophylaxis of infections in adult and pediatric patients receiving CAR T-cell therapy: a position paper. *Infection*. 2021;49(2):215–31. <https://doi.org/10.1007/s15010-020-01521-5>.
30. Hill JA, Seo SK. How I prevent infections in patients receiving CD19-targeted chimeric antigen receptor T cells for B-cell malignancies. *Blood*. 2020;136(8):925–35. <https://doi.org/10.1182/blood.2019004000>.
31. ASH. ASH-ASTCT COVID-19 Vaccination for HCT and CAR T Cell recipients: Frequently Asked Questions. 24/06/2021. [acessado em 4 julho 2021]. Disponível em: <https://www.hematology.org/covid-19/ash-astct-covid-19-vaccination-for-hct-and-car-t-cell-recipients>
32. Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA, et al. Axicabtagene ciloleucel CAR T-cell therapy in refractory large B-cell lymphoma. *N Engl J Med*. 2017;377(26):2531–44. <https://doi.org/10.1056/nejmoa1707447>.
33. Jain T, Knezevic A, Pennisi M, Chen Y, Ruiz JD, Purdon TJ, et al. Hematopoietic recovery in patients receiving chimeric antigen receptor T-cell therapy for hematologic malignancies. *Blood Adv*. 2020;4(15):3776–87. <https://doi.org/10.1182/bloodadvances.2020002509>.
34. Strati P, Varma A, Adkins S, Nastoupil LJ, Westin J, Hagemeister FB, et al. Hematopoietic recovery and immune reconstitution after axicabtagene ciloleucel in patients with large B-cell lymphoma. *Haematologica*. 2021;106(10):2667–72. <https://doi.org/10.3324/haematol.2020.254045>.
35. Baird JH, Epstein DJ, Tamaresis JS, Ehlinger Z, Spiegel JY, Craig J, et al. Immune reconstitution and infectious complications following axicabtagene ciloleucel therapy for large B-cell lymphoma. *Blood Adv*. 2021;5(1):143–55. <https://doi.org/10.1182/bloodadvances.2020002732>.
36. Locke FL, Ghobadi A, Jacobson CA, Miklos DB, Lekakis LJ, Oluwole OO, et al. Long-term safety and activity of axicabtagene ciloleucel in refractory large B-cell lymphoma (ZUMA-1): a single-arm, multicentre, phase 1–2 trial. *Lancet Oncol*. 2019;20(1):31–42. [https://doi.org/10.1016/S1470-2045\(18\)30864-7](https://doi.org/10.1016/S1470-2045(18)30864-7).
37. Hill JA, Krantz EM, Hay KA, Dasgupta S, Stevens-Ayers T, Bender Ignacio RA, et al. Durable preservation of antiviral antibodies after CD19-directed chimeric antigen receptor T-cell immunotherapy. *Blood Adv*. 2019;3(22):3590–601. <https://doi.org/10.1182/bloodadvances.2019000717>.
38. Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al. Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N Engl J Med*. 2018;378(5):439–48. <https://doi.org/10.1056/nejmoa1709866>.
39. Bhoj VG, Arhontoulis D, Wertheim G, Capobianchi J, Callahan CA, Ellebrecht CT, et al. Persistence of long-lived plasma cells and humoral immunity in individuals responding to CD19-directed CAR T-cell therapy. *Blood*. 2016;128(3):360–70. <https://doi.org/10.1182/blood-2016-01-694356>.
40. Walti CS, Krantz EM, Maalouf J, Boonyaratanakornkit J, Keane-Candib J, Joncas-Schronce L, et al. Antibodies against vaccine-preventable infections after CAR-T cell therapy for B cell malignancies. *JCI Insight*. 2021;6(11):e146743. <https://doi.org/10.1172/jci.insight.146743>.
41. Hill JA, Giral S, Torgerson TR, Lazarus HM. CAR-T – and a side order of IgG, to go? – Immunoglobulin replacement in patients receiving CAR-T cell therapy. *Blood Rev*. 2019;38:100596. <https://doi.org/10.1016/j.blre.2019.100596>.