



Original article

Prevalence of Torque teno virus in healthy donors of Paraná State, southern Brazil



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ABSTRACT

Objective: To determine the prevalence of the Torque teno virus in healthy donors in the northern and northwestern regions of the state of Paraná, southern Brazil.

Methods: The Torque teno virus was detected by a nested polymerase chain reaction using a set of oligoprimeres for the N22 region.

Results: The prevalence of the virus was 69% in 551 healthy blood donors in southern Brazil. There was no statistically significant difference between the presence of the virus and the variables gender, ethnicity and marital status. There was significant difference in the prevalence of the virus regarding the age of the donors (*p*-value = 0.024) with a higher incidence (74.7%) in 18- to 24-year-old donors.

Conclusion: A high prevalence of Torque teno virus was observed in the population studied. Further studies are needed to elucidate the routes of contamination and the clinical implications of the virus in the healthy population.

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Introduction

The Torque teno virus (TTV) was first detected in 1997 in the blood of Japanese patients with post-transfusion hepatitis.^{1,2} The virus was also detected in the liver and blood of people with hepatic pathologies of unknown etiology.² The association between TTV and liver diseases is still controversial,

and several studies have been undertaken to identify infection sources.³⁻⁵

Epidemiological studies have evidenced the prevalence of TTV in other pathological conditions, such as in autoimmune diseases,⁴ respiratory conditions⁶ and cancer.⁷ However, information is still lacking on TTV infection and the development of pathologies, as well as the change in the course of a particular disease.^{3,5}

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Table 1 – Prevalence of Torque teno virus in healthy populations of different Brazilian states.

Reference	Year of publication	State	Samples tested n	TTV DNA (%)
Niel et al. ¹⁷	1999	Rio de Janeiro	72	62.0
Bassit et al. ¹⁸	2002	São Paulo	197	85.3
de Castro Amarante et al. ¹⁹	2007	São Paulo	270	50.5
Pinto et al. ²⁰	2007	Pará	186	60.0
Nasser et al. ⁴	2009	Paraná	100	6.0
Costa et al. ⁵	2012	Mato Grosso do Sul	46	15.2
Massaú et al. ²¹	2012	Rio Grande do Sul	150	73.3

TTV: Torque teno virus.

Blood transfusion was initially indicated as the principal via of viral transmission due to direct contact with contaminated blood. Despite the progress in pretransfusion safety, blood recipients are not free from risk of contamination.⁸

The serological tests performed on blood donors in Brazil are established by the national health surveillance agency (ANVISA), and include serology for HIV1 and HIV2, HTLV1 and HTLVII, hepatitis B (HBV), hepatitis C (HCV), *Trypanosoma cruzi* (Chagas disease), *Treponema pallidum* (syphilis) and *Plasmodium* in endemic areas of malaria.⁹

Besides the serological tests conducted according to the Ministry of Health protocol, there is concern about the emerging and re-emerging diseases that can affect transfusion safety.¹⁰

However, new routes of transmission have been identified, due to the presence of the virus in different biological excretions such as in feces,¹¹ saliva¹² and also in river water contaminated by sewage.¹³

Currently, wide variability in the prevalence of TTV has been observed in healthy populations in different countries, such as in Alexandria in Egypt (48.4%),¹⁴ United Arab Emirates (75.0%)¹⁵ and Iran (13.4%).¹⁶ In Brazil the prevalence of TTV varies from 6 to 85% in different states (Table 1).

Several factors may contribute to the variability of the results of TTV prevalence studies, such as the geographical distribution of the population under analysis, the diagnostic method used, the size of the study group and the difficulty of making a single set of primers able to identify the majority of viral genotypes.^{10,15}

TTV infection is common in healthy donors worldwide.^{15,19} Knowledge of the prevalence of the TT virus in specific regions, serves as a resource to elucidate the transmission routes and the possible cause of disease, and may assist in developing guidelines for actions to control virus transmission in populations. The aim of this study was to determine the prevalence of TTV in healthy donors in the northern and northwestern regions of Paraná state, as there is a lack of studies showing the prevalence of the TTV virus in healthy donors in southern Brazil.

Methods

This transverse quantitative analysis involves human DNA samples obtained from healthy donors in 2010. The population comprised 551 volunteers, aged between 18 and 55 years. The samples were grouped according to the geographic location of

the source, in one of the seven mesoregions of Paraná State, according to the Brazilian Institute of Geography and Statistics (IBGE) (Northwest, Central-West, North-Central, the region of Norte Pioneiro, Central-Eastern, Mid-South and Metropolitan region of Curitiba) and other states.

A nested polymerase chain reaction (nested PCR) with specific primers for the N22 codifying region (ORF1 Open Reading Frame 1) was employed to detect TTV DNA. A sense primer (RD037) followed by oligonucleotide primers 5' GCA GCAGCA TAT GGA TAT GT 3' and RD038 (5' TGA CTG TGC TAA GGC CTC TA 3') were employed in the first amplification. The product of the first amplification and the antisense primers RD051 5' CAT ACA CAT GAA TGC CAG GC 3' and RD052 5' GTA CTT CTT GCT GGT GAA AT 3' were used in the second amplification. All reagents were identical in both reactions with a final volume of 25.0 µL comprised of 2.5 µL PCR buffer, 0.75 µL 50 mM magnesium chloride ($MgCl_2$), 2.0 µL 1.25 mM deoxyribonucleotide phosphate (dNTP), 1.0 µL of each sense (2.5 µM) and antisense (2.5 µM) primer, 2.5 µL Taq DNA polymerase (Invitrogen Life Technologies Brazil), and 12.75 µL sterile MilliQ water. A further 2.5 µL genomic DNA was used for the first amplification, and 2.5 µL of the product from the first amplification was used for the second amplification. Both reactions occurred in a thermocycler (Applied Biosystems) device with denaturing at 94 °C for 30 s, followed by 35 cycles at 53 °C for 30 s for primer annealing, 72 °C for 45 s for primer extension, with a final extension at 72 °C for 10 min. The amplified DNA products were analyzed by 2% agarose gel electrophoresis, stained with SYBR® Safe (1 µL/10 mL gel), and photographed under UV light. The DNA Ladder (Invitrogen) consisted of 50 base pairs (bp) and the amplified DNA products of 197 bp. All tests included a positive control (TTV genomic DNA). Data were analyzed with the Statistic 7.0 computer program using the chi-squared test, Yates's continuity correction and Fisher's exact test with significance set at a level of 5%. The assay complied with all ethical guidelines and was approved by the Research Ethics Committee of the Universidade Estadual de Maringá (UEM), Paraná, Brazil (Process 271/2011).

Results

The demographic data and the prevalence of TTV in healthy blood donors are shown in Table 2. The mean age of the donors was 33.7 years, and the majority were women 62.6% (345/551). Most of the donors were Caucasian 83.1% (458/551). TTV viral DNA was detected in 69% (380/551) of blood donors. Among the

Table 2 – Demographic characteristics and prevalence of Torque teno virus among healthy blood donors.

	TTV Positive n (%)	TTV Negative n (%)	Total	p-Value
Gender				0.785
Male	144 (70.0)	62 (30.0)	206	
Female	236 (68.4)	109 (31.6)	315	
Age (years)				0.024
18–24	124 (74.7)	42 (25.3)	166	
25–38	154 (70.3)	65 (29.7)	219	
39–53	97 (63.0)	57 (37.0)	154	
54–55	5 (41.7)	7 (58.3)	12	
Ethnic background				0.999
Caucasian	316 (69.0)	142 (31.0)	458	
African descent	64 (68.8)	29 (31.2)	93	
Marital status				0.916
Married	183 (68.0)	86 (32.0)	269	
Single	170 (69.7)	74 (30.3)	244	
Others ^a	27 (71.1)	11 (28.9)	38	

TTV: Torque teno virus.

^a Widowed, separated, divorced or cohabiting.

sociodemographic variables, the proportion of TTV-positive individuals differed in respect to age (Fisher's exact test: *p*-value = 0.024), with the rate being higher in the 18–24 year olds. There was no statistically significant difference in the prevalence of the virus between gender and ethnic background (*p*-value > 0.05).

The prevalence of TTV in healthy blood donors was assessed by mesoregion of the state of Paraná, southern Brazil (Table 3). The statistics test (Chi squared with Yates correction) indicated no significant differences in the presence of the virus and the different mesoregions (*p*-value = 0.576).

Discussion

The prevalence of TTV in blood donors in the mesoregions of the state of Paraná in southern Brazil was 69%. Other studies in Brazilian populations showed 60% prevalence in Belém, Pará²⁰ and 50.5% in the southeastern region of the state of São Paulo.¹⁸ In southern Brazil, studies showed a high prevalence of the virus in healthy donors (73.3%) in the municipality of Pelotas, RS²¹ and also the presence of the virus in samples of drinking water and sewage water.^{13,22}

TTV transmission by blood transfusion has been a recurring concern since the 1990s.¹ In Brazil, a study conducted at a university hospital showed concern about the risk of viral transmission by blood transfusion,²³ the serological screening of donors cannot provide complete protection from the transmission of infectious agents.

Similar to TTV, other viruses related to liver damage are overlooked in blood donors, including the hepatitis G virus (HGV). Some studies have shown the prevalence of HGV in healthy populations of Japan (0.9%) and South Africa (18.9%).²⁴ In Brazil a prevalence of 7.1% was also shown in the state of Goiás²⁵ and 9.7% in São Paulo.²⁶

Although TTV contamination can occur from both contaminated blood and blood products,² there is no specific

legislation that requires testing of blood donors for the virus. Therefore, little is known about the routes of transmission and diseases originating from the presence of the virus in the human population.

Based on the results presented, the association of the virus with the study variables can be determined (Table 2). The prevalence of TTV infection was (380/69%) in healthy donors from the northern and northwestern regions of Paraná, slightly below that found in Rio Grande do Sul (73.3%)²¹ and the region of São Paulo (85.3%).¹⁸ However, one should also note that the prevalence of the virus in other countries ranged from 2.7 to 79.5%.¹⁰ The variables of gender, race (Caucasian or African descent), and marital status showed no statistical association with the presence of the virus. The results of this study are in agreement with other studies that have suggested that TTV infection is relatively common in different populations and in different regions of the world.^{1,2}

With respect to age groups, the study included individuals between 18 and 55 years and revealed a high prevalence of infected young people between 18 and 24 (74.7%). However, it was found that the prevalence declined in over 24-year-old individuals, especially those of 54 and 55 (41.7%). This contrasts with previous studies that showed a cumulative prevalence with increasing age, or the presence of the virus independently of age.²⁷

Several studies conducted in different countries and individuals in different age groups demonstrated varying prevalences for TTV.^{18,28} The discussion of other age groups is limited by the particular population selected for this study. The results of this study indicated that the presence of TTV was significantly associated with age (*p*-value = 0.024), in agreement with a study in Pelotas, southern Brazil.²¹

The data for mesoregions (Table 3) indicate that there was no statistically significant difference between the presence of the virus and the samples from donors from different regions of Paraná. The grouping in mesoregions was necessary due to the large number of municipalities that comprised this study.

Table 3 – Distribution of the prevalence of Torque teno virus in healthy blood donors.

	TTV Positive n (%)	TTV Negative n (%)	Total n (%)	p-Value
Mesoregions				
North-West	169 (68.7)	77 (31.3)	246 (45.0)	0.576
Central-West	4 (80.0)	1 (20.0)	5 (0.9)	
North-Central	200 (68.5)	92 (31.5)	292 (53.0)	
Norte Pioneiro	1 (100.0)	0	1 (0.2)	
Central-East	0	1 (100.0)	1 (0.2)	
Central-South	2 (100.0)	0	2 (0.4)	
Metropolitan Curitiba	1 (100.0)	0	1 (0.2)	
Other States	3 (100.0)	0	3 (0.5)	
Total	380 (69.0)	171 (31.0)	551 (100.0)	

TTV: Torque teno virus.

Most of the donors were from the north-central and north-west Paraná mesoregions (98%) with prevalences of 68.5% and 68.7%, respectively. These regions belong to the 15th Regional Health District of Paraná and refer the city of Maringá, Paraná, Brazil. Abe et al. demonstrated that the TTV virus is widely distributed in different regions of the planet, and with high prevalence rates.²⁹

As is apparent from the literature, several factors may influence the variability of results for TTV prevalence, among them the geographical distribution of the populations studied, the diagnostic methods of detection, the size of study groups and the set of primers used in the study.^{3,15,28}

The high rates of viral prevalence may be directly related to the forms of contamination. A study of samples of blood and saliva from the same individuals showed the presence of the virus in the same proportions regardless of the biological sample used.³⁰ The presence of the virus in water has been investigated over time, and although the purpose of the present study was not to demonstrate the presence of virus in environmental samples, the importance of this analysis for studying the viral prevalence in a given region is important.

Studies have detected TTV in 97% of water samples collected in Japan³¹ and in Brazil, in 92% of samples collected from rivers and streams in Manaus.³² The viral genome was also reported in samples of drinking water in Rio Grande do Sul.¹³ The presence of TTV in the water of rivers, lakes, and treatment plants and especially in drinking water has had a major impact in spreading the virus. This may be related to the high prevalence of the virus in healthy individuals.

Conclusion

This study found a high prevalence of TTV in healthy blood donors, in agreement with other studies in the Brazilian population. The clinical significance of the presence of the virus in these donors cannot be evaluated based on this study, but can serve as a basis for future studies. In view of the different transmission routes and the lack of complete information about the pathogenesis of TTV, it is important to develop measures to minimize the risk of transmission of this and other viruses among healthcare providers.

Conflicts of interest

The authors declare no conflicts of interest.

REFERENCES

- Nishizawa T, Okamoto H, Konishi K, Yoshizawa H, Miyakawa Y, Mayumi M. A novel DNA virus (TTV) associated with elevated transaminase levels in posttransfusion hepatitis of unknown etiology. *Biochem Biophys Res Commun*. 1997;241(1):92–7.
- Okamoto H, Nishizawa T, Kato N, Ukita M, Ikeda H, Iizuka H, et al. Molecular cloning and characterization of a novel DNA virus (TTV) associated with posttransfusion hepatitis of unknown etiology. *Hepatol Res*. 1998;10(1):1–16.
- Watanabe MA, Miranda HC, Oliveira KB, Oliveira CE, Tristão FS, Duarte LM, et al. Aspectos patológicos, imunológicos e propriedades moleculares do TT vírus. *J Bras Patol Med Lab*. 2005;41(4):223–8.
- Nasser TF, Brajão de Oliveira K, Reiche EM, Amarante MK, Pelegrinelli Fungaro MH, Watanabe MA. Detection of TT virus in HIV-1 exposed but uninfected individuals and in HIV-1 infected patients and its influence on CD4+ lymphocytes and viral load. *Microb Pathog*. 2009;47(1):33–7.
- Costa MR, Costa IP, Devalle S, Castro ARCM, Freitas SZ. Prevalence and genetic diversity of torque teno virus in patients with systemic lupus erythematosus in a reference service in Mato Grosso do Sul. *Rev Bras Reumatol*. 2012;52(1):49–54.
- Maggi F, Pifferi M, Fornai C, Andreoli E, Tempestini E, Vatteroni M, et al. TT virus in the nasal secretion of children with acute respiratory diseases: relations to viremia and disease severity. *J Virol*. 2003;77(4):2418–25.
- Girard C, Ottomani L, Ducos J, Dereure O, Carles MJ, Guillot B. High prevalence of Torque Teno (TT) virus in classical Kaposi's sarcoma. *Acta Derm Venereol*. 2007;87(1):14–7.
- Carrazzone CF, Brito AM, Gomes YM. Importância da avaliação sorológica pré-transfusional em receptores de sangue. *Rev Bras Hematol Hemoter*. 2004;26(2):93–8.
- Ministério da Saúde. Agência Nacional de Vigilância Sanitária. Resolução da Diretoria Colegiada (RDC) No 153 de 14 de julho de 2004. Brasília: ANVISA; 2004.
- Karimi G, Gharehbaghian A, Tafti MF, Vafaiyan V. Emerging infectious threats to the blood supply: seroepidemiological studies in Iran – a Review. *Transfus Med Hemother*. 2013;40(3):210–7.

11. Okamoto H, Akahane Y, Ukita M, Fukuda M, Tsuda F, Miyakawa Y, et al. Fecal excretion of a nonenveloped DNA virus (TTV) associated with posttransfusion non-A-G hepatitis. *J Med Virol.* 1998;56(2):128-32.
12. Naganuma M, Tominaga N, Miyamura T, Soda A, Moriuchi M, Moriuchi H. TT virus prevalence, viral loads and genotypic variability in saliva from healthy Japanese children. *Acta Paediatr.* 2008;97(12):1686-90.
13. Vecchia AD, Fleck JD, Comerlato J, Kluge M, Bergamaschi B, Da Silva JV, et al. First description of Adenovirus, Enterovirus, Rotavirus and Torque teno virus in water samples collected from the Arroio Dilúvio, Porto Alegre, Brazil. *Braz J Biol.* 2012;72(2):323-9.
14. Hashish MH, El-Barawy MA, Mahmoud OA, Abdel Rahman NW. TT virus among blood donors in Alexandria. *J Egypt Public Health.* 2005;80(5-6):651-64.
15. Alfaresi MS, Elnazer AM, Alzaabi AS, Elkoush AA, Islam AA. Transfusion transmitted virus in screened United Arab Emirates blood donors. *Saudi Med J.* 2006;27(1):58-62.
16. Sara A, Solhjoo K, Jahromi AR, Yaghoobi R. Study the prevalence of TT virus infection in South Iranian volunteer blood donors. *Afr J Microbiol Res.* 2012;6(23):5077-81.
17. Niel C, de Oliveira JM, Ross RS, Gomes SA, Roggendorf M, Viazov S. High prevalence of TT virus infection in Brazilian blood donors. *J Med Virol.* 1999;57(3):259-63.
18. Bassit L, Takei K, Hoshino-Shimizu S, Nishiya AS, Sabino EC, Bassit RP, et al. New Prevalence Estimate of TT virus (TTV) infection in low- and high-risk population from São Paulo, Brazil. *Rev Inst Med Trop São Paulo.* 2002;44(4):233-4.
19. de Castro Amarante MF, Kashima S, Covas DT. TT virus (TTV) genotyping in blood donors and multiple transfused patients in Brazil. *Virus Genes.* 2007;35(3):503-9.
20. Pinto WV, Assis MF, Lemos JA. Prevalência do TTV em doadores de sangue, na região metropolitana de Belém-Pará. *Caderno Saúde Coletiva.* 2007;15(3):349-56.
21. Massaú A, Martins C, Nachtigal GC, Araujo AB, Rossetti ML, Niel C, et al. The high prevalence of Torque Teno Virus DNA in blood donors and haemodialysis patients in southern Brazil. *Mem Inst Oswaldo Cruz.* 2012;107(5):684-6.
22. Vecchia AD, Kluge M, dos Santos da Silva JV, Comerlato J, Rodrigues MT, Fleck JD, et al. Presence of Torque teno virus (TTV) in tap water in public schools from Southern Brazil. *Food Environ Virol.* 2013;5(1):41-5.
23. Schöninger N, Duro CL. Atuação do enfermeiro em serviço de hemoterapia. *Ciênc Cuid Saúde.* 2010;9(2):317-24.
24. Sathar MA, Soni PN, York D. GB Virus C/Hepatitis G Virus (GBV-C/HGV): still looking for a disease. *Int J Exp Pathol.* 2000;81(5):305-22.
25. Oliveira LA, Martins RM, Carneiro MA, Teles AS, Silva AS, Cardoso DD, et al. Prevalence and genotypes of GB Virus C/Hepatitis G virus among blood donors in Central Brazil. *Mem Inst Oswaldo Cruz.* 2002;97(7):953-7.
26. Levi JE, Contri DG, Lima LP, Takaoka DT, Garrini RH, Santos W, et al. High prevalence of GB Virus C/Hepatitis G Virus RNA among Brazilian blood donors. *Rev Inst Med Trop São Paulo.* 2003;45(2):75-8.
27. Hsieh SY, Wu YH, Ho YP, Tsao KC, Yeh CT, Lian YF. High prevalence of TT virus infection in healthy children and adults and in patients with liver disease in Taiwan. *J Clin Microbiol.* 1999;37(6):1829-31.
28. Vasconcelos HC, Menezes ME, Niel C. TT virus infection in children and adults who visited a general hospital in the south of Brazil for routine procedure. *Mem Inst Oswaldo Cruz.* 2001;96(4):519-22.
29. Abe K, Inami T, Asano K, Miyoshi C, Masaki N, Hayashi S, et al. TT virus infection is widespread in the general populations from different geographic regions. *J Clin Microbiol.* 1999;37(8):2703-5.
30. Spandole S, Cimponeriu D, Toma M, Radu I, Ion D. Rapid detection of human torque teno viruses using high-resolution melting analysis. *Balkan J Med Genet.* 2013;16(1):55-62.
31. Haramoto E, Katayama H, Oguma K, Yamashita H, Nakajima E, Ohgaki S. One-year monthly monitoring of Torque Teno Virus (TTV) in wastewater treatment plants in Japan. *Water Res.* 2005;39(10):2008-13.
32. Diniz-Mendes L, Paula VS, Luz SL, Niel C. High prevalence of human Torque Teno virus in streams crossing the city of Manaus, Brazilian Amazon. *J Appl Microbiol.* 2008;105(5):51-8.