Case Report

Compound heterozygous state of \( \beta \)-thalassemia with IVS1-5 (G→C) mutation and Indian deletion-inversion \( G\gamma(A\gamma\delta\beta)^0 \)-thalassemia in eastern India

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Introduction

It has recently been estimated that each year, more than seven million babies worldwide are born with either a congenital abnormality or a genetic disease.\(^1\) Hemoglobinopathies are the commonest autosomal hereditary disorders and present a major public health problem in India. The overall prevalence of the \( \beta \)-thalassemia trait is 2.78% but this varies from 1.48 to 3.64% in different states of India\(^2\) compared to the carrier frequency in Brazil (1%).\(^3\) Recently we reported that the carrier frequency of the \( \beta \)-thalassemia gene with the IVS1-5 (G→C) mutation in western districts of Odisha, India is 3.75%.\(^4,5\) In India, the IVS1-5 (G→C) mutation is the most common \( \beta \)-thalassemia mutation. However, the IVS1-5 (G→C) mutation along with other mutations, including IVS1-1 (G→T), Cd41/42 (-TCTT), Cd 8/9 and a 619 base pair deletion, accounts for >90% of mutations causing \( \beta \)-thalassemia. In Brazil, the Cd 39 (C→T) mutation is the most prevalent cause of \( \beta \)-thalassemia followed by the IVSI-6 (T→C), and IVSI-110 (G→A) mutations.\(^6\) A map showing zonal distribution of \( \beta \)-thalassemia mutations in India is depicted in Figure 1.

The Veer Surendra Sai Medical College and Hospital, Burla, Odisha is a tertiary care referral hospital catering for western Odisha as well as eastern districts of Chhattisgarh state. Under a comprehensive sickle cell care program, we screen outpatient and hospitalized cases in the Institute for Sickle Cell Disease and other hemoglobinopathies at the Sickle Cell Clinic and Molecular Biology Laboratory.

Compound heterozygotes for \( \beta \)-thalassemia and structural hemoglobin (Hb) variants usually present with a severe form of the disease. Here, we report a case, a compound heterozygote for the \( \beta \)-thalassemia mutation IVS1-5 (G→C) and Indian deletion-inversion \( G\gamma(A\gamma\delta\beta)^0 \)-thalassemia (HbVar ID-1038) from the district of Bargarh in the state of Odisha, India. The molecular structure of the Indian deletion-inversion \( [G\gamma(A\gamma\delta\beta)^0]^{-}-thal \) is caused by a major rearrangement.
An 18-year-old female patient (height 147 cm and weight 40 kg) belonging to the ‘Chasa’ caste was admitted to the Department of Medicine of Veer Surendra Sai Medical College and Hospital, Burla, Odisha, India with severe anemia, but with no history of any other medical complaints. Her Hb level was 2 g/dL at the time of admission to the hospital. After a transfusion of three blood units, the Hb level reached 7.8 g/dL. The erythrocyte sedimentation rate was 12 mm/h. Other parameters of blood indices after the blood transfusion were white blood cell count (WBC): 10.9 × 10^9 cells/L; red blood cells (RBC): 3.35 × 10^6/µL; hematocrit (HCT): 23.9%; mean corpuscular volume (MCV): 71.7 fl; mean cell hemoglobin (MCH): 20.6 pg; mean cell hemoglobin concentration (MCHC): 29.0 g/dL; and platelets (PLT): 149 × 10^3/L. Peripheral blood smear examination showed microcytic, hypochromic anemia, marked anisopelikocytes, many macrocytes, few microcytes and fragmented RBCs. Staining by the Kleihauer-Betke method demonstrated pancellular distribution of fetal hemoglobin (Hb F). Ultrasonography examination revealed hepatomegaly (16.5 cm) and splenomegaly (14.5 cm). An X-ray of chest (F-A view) and X-ray of both hip joints (A-P view) showed no abnormalities.

Automated high-performance liquid chromatography using the β-Thalassemia Short Program on Bio-Rad Variant-II system showed various fractions of Hb with a raised level of Hb F (84.6%) and an Hb A2 concentration of 6.9% (Figure 2). Parents’ studies revealed that her mother had a high Hb F level (13%) whereas her father had high Hb A2 level (5.2%). All hematological parameters of the case and her parents are shown in Table 1. Screening for the common molecular determinants of raised Hb F, the Indian deletion-inversion Gγ(Δγβ)β-thalassemia and hereditary persistence of fetal Hb (HPFH) was performed by gap-polymerase chain reaction (PCR) using primers and protocol as described previously.7

![Figure 1 - Map of India depicting the zonal distribution of β-thalassemia mutations.](image)
Common Indian β-thalassemia mutations were confirmed by multiplex amplification refractory mutation system (ARMS)-PCR using a previously described protocol. Moreover, alpha globin gene deletions (α−3.7 and α−4.2) were investigated by gap-PCR. The DNA study showed father as heterozygous for the IVS1-5(G→C) mutation and mother as a Gγ(Aγδβ)0-thalassemia carrier. The gel picture for Indian deletion-inversion Gγ(Aγδβ)0-thalassemia is shown in Figure 3. Alpha-thalassemia was not observed in any of them.

Written informed consent was obtained from the patient along with her parents and the study was approved by Institutional Ethics Committee of Veer Surendra Sai Medical College and Hospital, Burla, Odisha, India.

Discussion

β∗-Thalassemia is the second most common hemoglobinopathy in our clinic’s population. We have reported earlier the clinical and molecular characteristics of Gγ(Aγδβ)0-thalassemia in heterozygous as well as in compound heterozygous states with Hb S. In Hb S/Gγ(Aγδβ)0-thalassemia cases, the patients had repeated painful crises along with histories of blood transfusions. Indian deletion-inversion Gγ(Aγδβ)0-thalassemia in the heterozygous form has been reported in western India in the state of Maharashtra. Recently, Pandey et al. described seven cases with Indian deletion-inversion Gγ(Aγδβ)0-thalassemia, of which three cases were co-inherited with β-thalassemia with the IVS1-5(G→C) mutation. These three cases had low values of both Hb A2 (3.2%, 2.8%, and 2.7%) and Hb F (21.3%, 10.7% and 20.3%) compared to the 6.9% and 84.6% of Hb A2 and Hb F, respectively in the current case. Their three cases were transfusion dependent and showed moderate anemia. Our case is the first case of compound heterozygote state of β-thalassemia with IVS1-5(G→C) mutation and Gγ(Aγδβ)0-thalassemia with mild phenotype from eastern India. In this patient, there was no other probable cause to explain the acute episode of anemia in history or investigations. As parvovirus B19 is not investigated in our institution, this may well be a cause of the anemia in this patient. The milder clinical presentation is likely due to the association of a milder β∗-thalassemia allele with high
Hb F level (84.6%). Our observation of homogeneous F cell distribution and an elevated Hb F level in peripheral blood, is in agreement to the report of Waye et al. in an African-American with compound heterozygosity of a Black form of (Aγδβ)0-thalassemia and the −29 (A→G) β+ -thalassemia mutation.12 We only investigated two types of α-thalassemia (α-1.7 and α-4.2), the most prevalent in India. Another form of α-thalassemia may be a factor for milder clinical presentation of the patient.

The Indian subcontinent has a heterogeneous population with different hemoglobinopathies. These Hb disorders should be included in the prenatal diagnosis of patients with severe or mild thalassemia. The characterization of these hemoglobinopathies will facilitate a prevention and control program of hemoglobinopathies including thalassemia in this region.

**Conflicts of interest**

The authors declare no conflicts of interest

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**REFERENCES**